

Motivation

• The rapid spread of the COVID-19 has shown the importance for rapid and cost-effective testing of emerging new viruses. Diagnostics that can detect active infections are typically molecular based, gauging the presence of the pathogen. For example, the SARS-CoV-2 can be detected by various virology methods, most pre-eminently by the reverse transcriptase quantitative polymerase chain reaction (RT-PCR) which requires trained personnel, special equipment, and careful design of the primer and probe.



• To solve these problems, we proposed a quantum sensor based on Nitrogen-vacancy (NV) centers in nanodiamonds (NDs) that is capable to detect virus RNA, or any other RNA based stimulus. NV centers in diamond act as stable fluorescence markers and magnetic field sensors and have been investigated as quantum sensors for applications ranging from material science to chemistry and biology. A promising avenue to detect biological signals is to transduce them into magnetic noise - using, e.g., magnetic molecules, such as gadolinium (Gd) complexes. Then, like FRETbased biosensors, the presence of a stimulus (in this case the virus) is detected as it induces a change in the NV fluorescence, following its separation from the magnetic nanoparticle.



Ground State: Triple System

- Spin state can be optically polarized & readout at room temperature;
- NV centers in host nanodiamond (ND) with size down to \sim 20 nm;
- NDs are biocompatible and have high photo-stability;

Reference:

 Changhao Li, Rouhollah Soleyman, Mohammad Kohandel, and Paola Cappellaro, "SARS-CoV-2 Quantum Sensor Based on Nitrogen-Vacancy Centers in Diamond", Nano Lett. 2022, 22, 1, 43–49 and references in this article

Quantum Sensor Based on Spin Defects in Nanodiamonds

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Design/Simulation

(a) Development of an FND device for intracellular nucleic acids delivery and monitoring using nanodiamond particles that are coated with PEI. (b) Schematics of electrical charge density for FND-PEI and FND-PEI-DNA complexes and the corresponding band bending of energetic lev in the diamond. (c) The results for the intensity of oxidized FNDs and FND-PEI and FND-PEI-DNA complexes as a function of wavelength.



Meaning: the minimal detectable copies of virus RNA in one second

$$\sim \frac{1}{T_{1,bulk}} + \gamma_e^2 B_s^2 S_s(\omega) + \gamma_e^2 B_{Gd}^2 S_{Gd}(\omega)$$
$$\omega = \omega_0$$

$$C_{s,Gd} = (\frac{\mu_0 \gamma_e \hbar}{\pi})^2 C_{s,Gd} \pi \frac{n}{(d_0/2+l)^4}$$

$$_{d} = R_{Gd} / (R_{Gd}^{2} + \omega_{0}^{2})$$

$$r^2 \frac{n^{1/2}}{r_{min}^2} \Rightarrow R_{dip,Gd} \approx 21.1 \mathrm{ns}^{-1} \mathrm{nm} \times r^{-1}$$

/els	We coated PEI on ND
	particles and immobilized
	different ratio of DNA to

Sensor synthesis:

our ND particles and found the best DNA/ND ratio. Measuring Zeta potential shows the attachment and detachment of PEI and DNA to the ND surface.



Based of the Fluorescence resonance energy transfer (FRET) of NDs, we attached a Fl-dye to NDs to confirm RNA coupling and DNA detachment:

Next step: replace dye with Gd complex, and measure T_1



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esired Property	meaning	Our sensor
Sensitivity	low false negative	sensitivity >> RT-PCR
Specificity	low false positive	comparable with RT-PCR
Speed	return result as soon	< RT-PCR*
Quantitative	amount of virus	High sensitivity

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Fabrication/Results



Based of the Fluorescence resonance energy transfer (FRET) between NV center (donor) and dye molecule (acceptor)

ND	DOTA-N DNA-NH ₂	HS GdCl ₃ DNA-DOTA> DNA-	DOTA(Gd)		
	HO N N OH HO N N OH HO N N OH				
ie to FRET	DOTA-NHS	DNA-DOTA	DNA-DOTA(Gd)		
	Using NHS chemistry to couple aptamers to gadolinium-				
diral DNA	tetraazacyc	lododecane	tetraacetic		
	acid (DOTA-	Gd). DOTA-(ūd is a		
	common T1-weighted MRI contrast				
	agent, and its structure is shown				
. 5	here.				